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PII: S1538-7836(24)00780-3

DOI: <https://doi.org/10.1016/j.jtha.2024.12.031>

Reference: JTHA 913

To appear in: *Journal of Thrombosis and Haemostasis*

Received Date: 14 October 2024

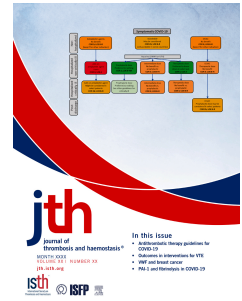
Revised Date: 26 December 2024

Accepted Date: 27 December 2024

Please cite this article as: Knauss EA, Guci J, Luc N, Disharoon D, Huang GH, Gupta AS, Nieman MT, Mice with Reduced PAR4 Reactivity show Decreased Venous Thrombosis and Platelet Procoagulant Activity, *Journal of Thrombosis and Haemostasis* (2025), doi: <https://doi.org/10.1016/j.jtha.2024.12.031>.

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1 **Mice with Reduced PAR4 Reactivity show Decreased Venous Thrombosis and Platelet**
2 **Procoagulant Activity**

3

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12 Running Title: PAR4-P322L decreases venous thrombosis in mice

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24 Abstract word count: 221

25 Total word count: 4759

26 Total figure count: 7

27

28

29

1 Abstract**2 Background**

3 Hypercoagulation and thrombin generation are major risk factors for venous thrombosis. Sustained
4 thrombin signaling through PAR4 promotes platelet activation, phosphatidylserine exposure, and
5 subsequent thrombin generation. A single-nucleotide polymorphism in PAR4 (rs2227376) changes
6 proline to leucine extracellular loop 3 (P310L), which decreases PAR4 reactivity and is associated with
7 a lower risk for venous thromboembolism (VTE) in a GWAS meta-analysis.

9 Objective

10 The goal of this study is to determine the mechanism for the association of rs2227376 with reduced risk
11 for VTE in using mice with a homologous mutation (PAR4-P322L).

13 Methods

14 Venous thrombosis was examined using our recently generated PAR4-P322L mice in the inferior vena
15 cava stasis and stenosis models. Coagulation and clot stability was measured using rotational
16 thromboelastometry (ROTEM). Thrombin generating potential was measured in platelet-rich plasma.
17 Phosphatidylserine surface expression and platelet-neutrophil aggregates were analyzed using flow
18 cytometry.

20 Results

21 PAR4^{P/L} and PAR4^{L/L} had reduced size of venous clots at 48 hours. PAR4^{P/L} and PAR4^{L/L} platelets had
22 progressively decreased phosphatidylserine in response to thrombin and convulxin, in addition to
23 decreased thrombin generation and decreased PAR4-mediated platelet-neutrophil aggregation.

25 Conclusions

26 The leucine allele in extracellular loop 3, PAR4-322L leads to fewer procoagulant platelets, decreased
27 endogenous thrombin potential and reduced platelet-neutrophil aggregation. This decreased ability to
28 generate thrombin and bind to neutrophils offers a mechanism for PAR4's role in VTE highlighting a key
29 role for PAR4 signaling.

31 Keywords:

32 protease-activated receptor 4, blood platelets, thrombin receptor, single nucleotide polymorphisms,
33 animal model

34

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1 Introduction

2 Venous thrombosis is driven by three factors known as Virchow's triad: blood flow stasis, endothelial
3 activation, and hypercoagulation [1]. Clot development involves an interplay between immune cells,
4 platelets, endothelial cells, and coagulation factors in a process known as immunothrombosis [2,3].
5 Neutrophils and platelets bind to activated endothelial cells and express a procoagulant surface that
6 promotes coagulation and thrombin generation [4–6]. Thrombin is produced in massive amounts at the
7 site of clot development via the coagulation cascade [7,8]. Its main functions are to convert fibrinogen
8 to fibrin, an essential component of venous thrombi [9]. Thrombin also activates cells, predominantly
9 endothelial cells and platelets, directly via protease-activated receptors (PARs) [10]. On human
10 platelets, thrombin cleaves PAR1 and PAR4 [11]. PAR1 elicits a rapid transient signaling cascade,
11 whereas PAR4 leads to prolonged signaling and is required for sustained platelet activation [12]. PAR4
12 requires more thrombin for activation than PAR1, and its unique signaling kinetics make it a promising
13 therapeutic target to prevent excessive and pathological platelet activation. PAR4 cleavage by thrombin
14 leads to platelet activation and aggregation [13]. PAR4 antagonists, like BMS-986120 and BMS-
15 986141, show significant potential in animal models and early clinical trials with a low risk of bleeding
16 and a high antithrombotic potential [14–16]. Given that PAR4-mediated platelet activation also
17 promotes venous thrombosis in mice [17], it is worth further exploring PAR4 as a therapeutic target
18 against platelet activation in thrombosis and inflammation.

19
20 Platelets are widely known for their essential roles in hemostasis and preventing excessive blood loss
21 upon injury [18]. Platelets respond to vascular damage and inflammation by adhering to the activated
22 endothelium and aggregating with each other [19]. After sustained activation they develop a
23 procoagulant surface covered in phosphatidylserine that serves as a scaffold for the assembly of
24 coagulation factors, leading to thrombin and fibrin generation to plug the broken vasculature [4].
25 Unchecked coagulation and excessive platelet activation leads to thrombosis. Platelets are known to
26 promote arterial thrombosis and have been targeted for prevention [20]. Platelets also drive the early
27 stages of venous thrombosis, but antiplatelets are not currently used to prevent venous events. Current
28 preventative therapies for venous thrombosis and its complication pulmonary embolism, known
29 collectively as venous thromboembolism (VTE), target coagulation to shut down thrombin generation
30 and fibrin production. The downside of this approach is that normal hemostasis is also impaired,
31 leading to bleeding complications. A targeted approach that reduces PAR4 activity would spare
32 thrombin's function on fibrinogen and other receptors like PAR1, allowing hemostasis to work
33 unhindered.

34
35 Single nucleotide polymorphisms (SNPs) in PAR4 impact signaling and subsequent platelet function
36 [21]. These SNPs impact patient response to antiplatelet therapies, and they can also be utilized as a
37 tool in laboratories to further investigate the therapeutic potential of reducing PAR4 reactivity [22,23]. A
38 SNP in PAR4 results in an amino acid switch at position 310 from a proline to a leucine (PAR4-P310L,
39 rs2227376) [23]. The leucine at 310 is associated with reduced PAR4 signaling in cells, as measured
40 by calcium signaling [23]. To determine how the PAR4-P310L variant impacts PAR4 function *in vivo*,
41 we developed a mouse model with a homologous mutation at position 322 (PAR4-P322L). Mice with
42 one (PAR4-322^{P/L}, heterozygous) or two (PAR4-322^{L/L}, homozygous) leucine alleles had a reduced
43 platelet response to the PAR4-agonist peptide and thrombin. These mice also showed impaired arterial
44 thrombosis and decreased clot stability [24].

45
46 Our lab, in collaboration with the INVENT consortium, also found that the leucine allele at PAR4-310 is
47 associated with a 15% risk reduction in VTE [23]. Here, we used our mouse model to investigate the

1 mechanism behind this association between PAR4 activity and venous thrombosis. PAR4-322^{P/L} and
2 PAR4-322^{L/L} mice had significantly reduced venous thrombosis, and mice without PAR4 (PAR4^{-/-}),
3 were less likely to develop venous thrombi in the early stages. PAR4-P322L also resulted in decreased
4 platelet procoagulant activity. We also showed that decreased PAR4 reactivity directly reduces platelet-
5 neutrophil aggregation. These data demonstrate that PAR4 drives immunothrombosis by promoting
6 platelet-neutrophil binding and inducing a procoagulant platelet phenotype that supports thrombin
7 generation. Importantly, the PAR4-P322L mutation did not significantly impact hemostatic clot
8 development as measured by rotational thromboelastometry, suggesting that PAR4 plays a more
9 significant role in pathological platelet activation.

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1 **Methods**

2 Animals

3 All animal experiments were performed in accordance with the approval from the Case Western
4 Reserve University (CWRU) Animal Ethics Committee. The PAR4-P322L mutation was introduced into
5 F2r13 by inducing a C to T nucleotide switch using the CRISPR-Cas9 genome-editing system, as we
6 have previously described [24]. PAR4 knockout mice (F2r13^{-/-}) were purchased from Mutant Mouse
7 Regional Resource Centers. These mice are referred to as PAR4-322^{P/P} (wild type), PAR4-322^{P/L},
8 PAR4-322^{L/L}, or PAR4^{-/-}. PAR4-P322L mice were bred via heterozygous crossings and genotyped via
9 sanger sequencing. Male and female mice were used for all *ex vivo* experiments.

11 Venous Thrombosis Models

12 Venous thrombosis was modeled in male mice using the stasis and stenosis models of inferior vena
13 cava (IVC) ligation [25,26]. For both models, mice were anesthetized using isoflurane (2%) and sterile
14 technique was performed to open the abdomen and isolate the IVC. Visible back branches were
15 cauterized, and the side branch was completely ligated when located below the renal vein. For the
16 stasis model, the IVC was completely ligated with nylon suture (7-0, Ethilon). For the stenosis model,
17 the suture was tied around a spacer (30g needle), which was removed immediately after ligation. For
18 both surgeries, the peritoneum and outside skin were closed separately with nylon sutures, and 500-
19 700 μ L of saline was injected subcutaneously. Mice were subcutaneously given buprenorphine (0.05
20 mg/kg) immediately following surgery, then every 8-12 hours for pain management. Mice were placed
21 on a heating pad daily and monitored for up to 48 hours.

23 Clot Staining

24 Clots were harvested from mice at 48 hours and dissected from the surrounding tissue. Clots were
25 added to tissue cassettes then fixed in 4% paraformaldehyde for 24 hours and stored in 70% ethanol at
26 4°C until processed. Fixed clots were embedded in paraffin and cut in 5 μ M sections by the Tissue
27 Resource Core at CWRU. H&E staining was also done by the tissue resource core at CWRU. After
28 H&E staining, tissue blocks were sent to the imaging core at the Cleveland Clinic Lerner Research
29 Institute for Carstairs staining.

31 Evaluation of Blood Coagulation using ROTEM

32 Rotational thromboelastometry (ROTEM) was used to assess blood coagulation in whole blood from
33 mice. 750 μ L of whole blood was collected via IVC puncture into 10% sodium citrate (80 μ L; 0.109 M).
34 ROTEM was performed on a quad-channel computerized device (ROTEM Delta TEM International,
35 Munich, Germany). Extrinsic pathway activation was assessed using the EXTEM assay. Within 2 hours
36 of blood collection, 300 μ L of each sample was placed in a disposable cuvette with an electric
37 automated pipette connected to the ROTEM system. 20 μ L of 0.2 M CaCl₂ was added to recalcify the
38 samples, followed by the addition of the EXTEM substrate (tissue factor) immediately before
39 measurements began. The assay was run for 60 minutes. Clotting time (CT), clot formation time (CFT),
40 maximum clot firmness (MCF), alpha angle, and the amplitudes at 10 (A10) and 20 (A20) minutes were
41 determined [27,28]. CT is the time from the initiation of the reaction until the amplitude reaches 2 mm.
42 CFT is the time between 2 and 20 mm of clot firmness. Alpha angle is the angle between the baseline
43 and the clotting curve at 2 mm. MCF refers to the maximum amplitude of clot firmness. A10 and A20
44 represent the clot amplitude at 10 and 20 minutes, respectively.

1 Thrombin Generation

2 Endogenous thrombin generating potential in platelet-rich plasma (PRP) was assessed using the
3 TechnoThrombin® thrombin generation assay (TGA) kit (Diapharma, Cat #5006010). Whole blood was
4 collected from mice via IVC puncture into sodium citrate. PRP from wild-type (PAR4-322P/P), PAR4-
5 322^{P/L}, PAR4-322^{L/L} or PAR4^{-/-} mice was prepared by centrifuging whole blood at 2300xg for 20
6 seconds. The plasma supernatant was carefully removed without disturbing the lower layer of red blood
7 cells. Platelets were counted using a Coulter Counter (Beckman Coulter), and PRP was diluted to
8 2×10^8 platelets/mL and added to a 96-well plate. TechnoThrombin® Reagent C High was used to
9 initiate the reaction and a fluorogenic substrate was used to monitor thrombin generation at 37°C for
10 60 minutes. Fluorescence was measured immediately after addition of the reagent/substrate mixture
11 with a Flexstation 3 (Molecular Devices) with SoftMax Pro software. The plate was read every minute
12 for one hour at a 360 nm excitation and 460 nm emittance. A standard concentration of thrombin was
13 diluted and measured over 10 minutes to create a standard curve for data analysis. Control samples
14 containing high and low concentrations of human plasma were included to show daily variation in
15 sample preparation and data collection.

16

17 Procoagulant Platelet Assay

18 Phosphatidylserine (PS) exposure was detected on washed platelets using flow cytometry for FITC-
19 tagged bovine lactadherin (Prolytix #BLAC-FITC). Whole blood was collected via IVC puncture into
20 sodium citrate (0.109 M), diluted 1:4 in platelet wash buffer (10 mM sodium citrate, 150 mM NaCl, 1
21 mM EDTA, 1% dextrose) supplemented with PGI₂, and centrifuged at 300xg for 5 minutes. The PRP
22 supernatant was collected, diluted 4:1 in platelet wash buffer with PGI₂, and centrifuged at 3000xg for
23 10 minutes. Pelleted platelets were resuspended in HEPES tyrode's buffer (140 mM NaCl, 6.1 mM KCl,
24 2.4 mM MgSO₄·7H₂O, 1.7 mM Na₂HPO₄, 5.8 mM Na-HEPES), counted, then diluted to 1×10^8
25 platelets/mL. 20 µL of diluted platelets were incubated with 5 µL convulxin, and 5 µL of either thrombin
26 (Prolytix, BCT-1020) or PAR4-agonist peptide (AYPGKF). The final concentration of agonist in each
27 reaction was 1 nM convulxin and 200 µM AYPGKF, 1 nM thrombin, or 5 nM thrombin. Following
28 activation, platelets were labeled with 12 µg/mL of APC-tagged CD41a (eBioMWRReg30, invitrogen #17-
29 0411-80) and 5 µg/mL FITC-Lactadherin for 15 minutes in the dark. Samples were fixed with 2%
30 paraformaldehyde and analyzed on a BD LSRII flow cytometer. Stimulation with a calcium ionophore
31 (A23187, 15 µM) was included for each sample to define the positive population (data not shown).

32

33 Platelet-Neutrophil Aggregates

34 Platelet-neutrophil aggregates were isolated as described previously [29]. Whole blood was collected
35 via IVC puncture and diluted 1:10 in M199 supplemented with 100 U/mL heparin. White blood cells
36 were labeled with PE-CD45 (clone 30-F11, invitrogen #12-0451-82), neutrophils with FITC-Ly6G (clone
37 1A8, invitrogen #11-9668-82), and platelets with APC-CD41a (eBioMWRReg30, invitrogen #17-0411-80).
38 Diluted whole blood was labeled and activated for 15 minutes at 37°C in the dark. BD FACs lysing
39 solution was added for 15 minutes to lyse red blood cells and fix the remaining cells. Samples were
40 centrifuged at 800xg for 5 minutes, and the pelleted cells were resuspended in phosphate-buffered
41 saline. Platelet neutrophil-aggregates were analyzed on a BD LSRII flow cytometer at the Case
42 Cytometry and Imaging Microscopy core.

Results

PAR4-P322L reduces venous thrombosis.

The single nucleotide polymorphism at rs2227376 results in a C to T base substitution leading to an amino change from proline to leucine at position 310 in extracellular loop 3 of PAR4. Individuals with the minor allele, leucine, have a 15% risk reduction in VTE. To study the impact of this substitution, we generated a mouse model with the homologous mutation, PAR4-P322L [24]. These mutant mice display impaired platelet activation and aggregation, as well as delayed arterial thrombosis [24].

To determine whether PAR4-P322L impacts venous thrombosis, our initial experiments used the complete stasis model of inferior vena cava (IVC) ligation. PAR4-322^{P/P} (wild type) mice develop fully formed thrombi by 48 hours following complete IVC ligation. The IVC containing the thrombus was harvested and weighed (**Figure 1A**). The average weight of IVCs without thrombi (13.6 mg, data not shown) was subtracted to give the weight of the thrombus alone. Thrombi from PAR4-322^{P/P} mice averaged 17.7 ± 4.2 mg. Thrombi from PAR4-322^{P/L} mice were 8.4 ± 5.1 mg ($p=0.004$), while the sizes of clots from PAR4-322^{L/L} mice had a wide range that averaged 12.5 ± 8.5 mg. Thrombi from PAR4 knockout mice (PAR4^{-/-}) were 8.1 ± 4.5 mg, significantly smaller than wild-types ($p=0.002$).

Platelets have critical roles in the early stages of venous clot development [30]. To determine the impact of PAR4 at the initiation phase, we examined the time course of thrombus growth in PAR4-322^{P/P} mice at 12, 16, 20, and 24 hours. At 12, 16, and 20 hours, small clots formed at an incidence of $\leq 50\%$ (data not shown). By 24 hours, PAR4-322^{P/P} mice developed both large and small thrombi more than half of the time (71% clot incidence) (**Figure 1B**). PAR4-322^{P/L} and 322^{L/L} mice similarly developed clots at 80% and 57% incidence, respectively. PAR4^{-/-} mice, however, were unable to develop clots by 24 hours. The thrombi that developed by 24 hours were 15.9 ± 10.1 mg for PAR4-322^{P/P}, 12 ± 6 mg for PAR4-322^{P/L}, and 10.7 ± 5.6 mg for PAR4-322^{L/L} (**Figure 1C**). Altogether, reducing or removing PAR4 activity decreased IVC thrombosis induced by complete venous stasis.

Partial stenosis of the IVC maintains residual blood flow and triggers inflammation at the site of clot development [26,31]. A major difference is the variability in resulting thrombus size and incidence compared to the stasis model. The reported variation is 65-100% thrombus occurrence at 48 hours. Wild-type mice developed clots 73% of the time (**Figure 2**). 67% of PAR4-322^{L/L} mice and 40% of 322^{P/L} mice developed thrombi by 48 hours. The clots that formed in PAR4-322^{P/P} (WT) mice were 21.0 ± 6.2 mg (**Figure 2**). Those from PAR4-322^{P/L} and PAR4-322^{L/L} mice were 8.8 ± 4.5 mg and 6.2 ± 4.8 mg ($p=0.01$), respectively. Overall, reducing PAR4 reactivity had an even greater impact on IVC stenosis than complete stasis.

Clots that developed after IVC stenosis were processed for H&E and carstairs staining to observe any differences in clot composition or fibrin deposition, respectively. Both the PAR4-P322L mice and wild-types developed clots with layers of fibrin and white blood cells as well as lines of platelets [30,32]. These characteristics are similar to those that define human thrombi, and indicate features that are essential to forming and stabilizing venous clots. H&E staining showed that while the clots from PAR4-322^{P/L} and PAR4-322^{L/L} mice were noticeably smaller than from wild-types, their composition was comparable (**Figure 3 panels A and B**). Thrombi from PAR4-P322L mice also showed a similar morphology to wild-types after carstairs staining (**Figure 3 panels C and D**). Thus, while the size of the clots decreases with PAR4 reactivity, the overall morphology and fibrin deposition remains the same.

Lack of PAR4 delays clot formation in ROTEM

We next used rotational thromboelastometry (ROTEM) to investigate the clot stability in whole blood from PAR4-322^{P/P}, PAR4-322^{P/L}, PAR4-322^{L/L}, and PAR4^{-/-} mice. While ROTEM is traditionally used to measure hemostatic function, here we are specifically using it to measure tissue factor-induced coagulation using EXTEM. Thromboelastograms were developed for each sample over 60 minutes and clotting time (CT), clot formation time (CFT), maximum clot firmness (MCF) were determined from these traces (**Figure 4**). Clotting time (CT), also known as the coagulation time, indicates the initial activation of the coagulation cascade via tissue factor (**Figure 4B**) [27]. CT was unchanged in blood from mutant PAR4-P322L mice and global PAR4^{-/-} mice in comparison to wild-types, averaging around 40 seconds for each genotype. Clot formation time (CFT) was also unchanged in blood from both PAR4-322^{P/L} and 322^{L/L} mice (**Figure 4C**). CFT reflects the kinetics of clot formation and predominantly depends on platelet function, thrombin generation, and fibrin accumulation [27]. Blood from PAR4^{-/-} mice showed a pronounced increase in CFT and averaged 373 ± 93.9 seconds compared to wild-types at 41 ± 2.7 seconds (p < 0.0001). The MCF was unchanged across genotypes indicating that PAR4 does not impact the overall strength of the clot (**Figure 4D**) [27]. The rate of fibrin formation (alpha angle), and clot firmness (amplitude) at early time points were not different between PAR4-322^{P/P}, PAR4-322^{P/L}, PAR4-322^{L/L} mice, however all decreased in PAR4^{-/-} mice (**Figure 4E-G**). Altogether, the allele at position 322 did not impact clot development in ROTEM stimulated with tissue factor. However the absence of PAR4 significantly delayed clot formation.

PAR4-P322L decreases phosphatidylserine exposure and thrombin generation.

Thrombin is produced in massive amounts at the site of venous thrombosis where it activates platelets via PAR4 and cleaves fibrin to crosslink and stabilize venous clots [33]. At high concentrations of strong agonists like thrombin and collagen, platelets will adopt a procoagulant phenotype that serves as a platform for thrombin generation [5] [34,35]. We tested thrombin generating potential of platelet-rich plasma (PRP) from PAR4-322^{P/P}, PAR4-322^{P/L}, PAR4-322^{L/L} and PAR4^{-/-} mice for 60 minutes. (**Figure 5A**). The total thrombin generated (area under the curve), peak thrombin, and time to peak thrombin all showed a similar downward trend that correlated with PAR4 reactivity (**Figure 5B-D**). All parameters were significantly decreased in PRP from PAR4^{-/-} mice. Taken together, thrombin generating potential concomitantly decreased as PAR4 reactivity decreased.

Platelet-mediated thrombin generation requires a procoagulant cell surface coated with phosphatidylserine (PS). We used FITC-labeled lactadherin to measure PS exposure on washed platelets. Platelets were stimulated with convulxin in combination with thrombin or the PAR4-agonist peptide (PAR4-AP), AYPGKF (**Figure 6**). When stimulated with convulxin (1 nM) combined with AYPGKF (200 μM), 7.4% of PAR4-322^{P/P} platelets were PS-positive compared to 4.8% for PAR4-322^{P/L} platelets. Platelets from PAR4-322^{L/L} and of PAR4^{-/-} mice did not respond at this dose. Platelets from both PAR4-322^{P/L} and 322^{L/L} had less PS exposure on the surface following stimulation with convulxin and 1 or 5 nM thrombin. The PAR4^{-/-} platelets did not respond to thrombin as expected. These data indicate that the leucine allele at position 322 negatively impacts platelet PS exposure and subsequent thrombin generation, a phenotype that is amplified in platelets deficient in PAR4.

PAR4-P322L decreases platelet-neutrophil complex formation.

PAR4 activation is known to induce surface P-selectin exposure, integrin activation, and platelet-neutrophil binding [36]. Additionally, procoagulant platelets preferentially activate neutrophils and promote thromboinflammation [5,37]. We have shown that mutant PAR4-P322L platelets have impaired

1 P-selectin exposure and integrin activation in response to the PAR4-AP and thrombin [24]. Since
2 platelet-neutrophil complexes (PNCs) are a risk factor for VTE, we examined the impact of PAR4-
3 P322L on platelet-neutrophil aggregation. PNCs were defined as the percentage of CD45+, Ly6G+
4 cells that were positive for CD41. Complexes were measured at increasing concentrations of AYPGKF
5 (50-400 μ M) (**Figure 7A-D**). PAR4-322^{P/P} showed an increase in PNCs in response to 50 μ M AYPGKF,
6 while PAR4-P322L and PAR4^{-/-} did not respond (**Figure 7B**). PAR4-322^{P/L} mice responded to 200 μ M,
7 but the percentage of PNCs were significantly less than PAR4-322^{P/P} (**Figure 7C**). In contrast, PAR4^{L/L}
8 and PAR4^{-/-} mice did not respond at the highest dose tested (400 μ M) (**Figure 7D**). These data
9 demonstrate that PAR4 reactivity is directly correlated to the formation of PNCs.

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1 Discussion

2 The single-nucleotide polymorphism (SNP) that results in PAR4-Pro310Leu (rs2227376) lowers PAR4
3 activity and decreases PAR4-mediated platelet activation and arterial thrombosis [24]. The leucine
4 allele at PAR4-310 is also associated with a 15% risk reduction in venous thromboembolism (VTE) [23].
5 Here, we used our mouse model of the PAR4-P310L variant (PAR4-P322L) to investigate the role of
6 PAR4 in venous thrombosis. The PAR4-P322L mouse model has a point mutation at a site homologous
7 to human PAR4-P310 and offers a way to study reduced PAR4 function *in vivo*. Here, we have shown
8 that PAR4-P322L decreases platelet procoagulant activity and negatively impacts venous thrombosis.
9

10 We used the stasis model of inferior vena cava (IVC) thrombosis to mimic a completely occlusive
11 thrombi in PAR4-322^{P/P} (wild-type), PAR4-322^{P/L}, 322^{L/L}, and PAR4^{-/-} mice. Complete IVC ligation is
12 known for consistently inducing large thrombi with a low variability in size, making it a reliable technique
13 for modeling acute thrombosis [31,38]. The PAR4-P322L mutation reduces PAR4 reactivity in response
14 to thrombin [24]. These data suggest it also reduce platelet and endothelial cell activation in the early
15 stages of thrombosis leading to smaller thrombi. By 48 hours, when the clots had fully formed, both
16 PAR4-322^{P/L} and PAR4^{-/-} mice had significantly smaller venous thrombi than wild-types. Interestingly,
17 PAR4-322^{L/L} mice developed thrombi with a much greater variability in size, with some as large as wild-
18 types and others as small as PAR4^{-/-} (**Figure 1A**). When clots were harvested at 24 hours, PAR4^{-/-}
19 were unable to develop venous thrombi, and PAR4-322^{L/L} mice only formed clots 50% of the time
20 (**Figure 1B**). This suggests that PAR4 is driving the early stages of VTE when immune cells interact
21 with coagulation factors to generate fibrin and initiate thrombosis [30].
22

23 Venous thrombi that develop from the stasis model of IVC ligation are similar in structure to those from
24 humans, however they do not arise in the same manner. Complete IVC stasis predominantly produces
25 clots through endothelial damage and tissue factor (TF) activation. The stenosis model of partial IVC
26 ligation produces clots through a mechanism that is more dependent on immune cell activation [31].
27 The stenosis model avoids extreme endothelial damage, and mimics less occlusive thrombi that are
28 more commonly seen in humans [26]. A downside of this model is that thrombosis is less likely to
29 occur, but it allows a more sensitive method for analyzing thrombosis incidence. Indeed, we saw that
30 both PAR4-322^{P/L} and PAR4-322^{L/L} mice developed fewer venous thrombi at 48 hours after partial IVC
31 ligation (**Figure 2**). Additionally, the clots that developed in the mutant mice were significantly smaller
32 than wild-types (**Figure 2**). The difference between the PAR4-P322L and wild-type mice was more
33 pronounced in the stenosis model compared to complete stasis. The loss of PAR4 activity showed the
34 greatest impact in the stasis model at early time points, suggesting a critical role in immunothrombosis
35 and clot initiation. Unlike complete stasis, partial stenosis retains residual blood flow and triggers clot
36 development through local inflammation and immunothrombosis [26]. Given that PAR4 is most highly
37 expressed in this system on platelets and endothelial cells, it is likely that these cells are mediating
38 PAR4's role in thrombosis. Additionally, our data is consistent with others that showed platelet-specific
39 knockout mice had decreased venous thrombus size and incidence after IVC stenosis [17]. Knowing
40 that the PAR4-P322L mutation directly impacts platelet activation, we used our mouse model to further
41 explore PAR4-mediated platelet activation in venous thrombosis.
42

43 Venous clots initiated by complete ligation of the inferior vena cava are heavily driven by TF exposed
44 upon endothelial damage [25]. The exact mechanism and source of TF is still unclear [39]. The lack of
45 PAR4 increased clot formation time (CFT) in ROTEM initiated by TF (EXTEM) (**Figure 4C**). The
46 increased clot formation time in the PAR4^{-/-} mice suggests PAR4 promotes the early steps of clot
47 formation but does not drive initiation of coagulation, overall clot stability, or clot firmness in this assay.

1 In contrast, the clot formation time in blood from PAR4-P322L mice was identical to wild-type mice. It is
2 not surprising that EXTEM does not distinguish between the PAR4 genotypes. It uses a strong activator
3 of the coagulation system (TF) and there are multiple signaling pathways independent of PAR4 that
4 lead to integrin activation and clot stability. In contrast, *in vivo* assays reveal an essential role for PAR4.
5 Lee et al. used the saphenous vein bleeding assay to show PAR4 on platelets is critical to fill the
6 puncture in the vessel wall [17]. Similarly, we have previously shown that the PAR4-P322L mutation
7 prolongs bleeding in the tail vein assay, albeit to a lesser extent than PAR4 knockout mice [24]. Even
8 with the blunt nature of ROTEM, we observed a sharp decrease in the alpha angle, which further
9 supports a role for PAR4 in thrombin generation. Since procoagulant platelets are essential for
10 mediating thrombin generation in hemostasis and thrombosis, impaired thrombin generation may
11 contribute to the mechanism for the decreased size and incidence of venous thrombi in our mutant
12 mice in addition to platelet aggregation.

13
14 Platelets promote coagulation by directly supporting thrombin activation and fibrin generation [7]. When
15 platelets are exposed to a high concentration of agonist(s), as in vascular damage or thrombosis, they
16 express phosphatidylserine on their surface [4,5]. This surface acts as a scaffold for the
17 prothrombinase complex to assemble and generate thrombin [6,40]. Thrombin's major function is to
18 mediate fibrinogen conversion to fibrin, an insoluble protein that aggregates and stabilizes blood clots
19 [19,41]. Thrombin also activates cells involved in thrombosis, including platelets, through protease
20 activated receptors (PARs) [10]. When platelets are simultaneously activated *ex vivo* with a PAR4
21 agonist and a GPVI agonist, they develop a procoagulant phenotype expressing phosphatidylserine on
22 their membrane [4]. Here, we showed that platelets from PAR4-322^{L/L} and PAR4^{-/-} mice had significantly
23 less phosphatidylserine expression after simultaneous stimulation of PAR4 and GPVI (**Figure 5**).
24 Platelet-mediated thrombin generation also decreased as PAR4 reactivity decreased (**Figure 4A-C**).
25 Altogether, we showed that as PAR4 reactivity decreases, platelet procoagulant activity also
26 decreases. Procoagulant platelet activation is known to promote arterial and venous inflammation and
27 thrombosis. This, along with platelet adhesion and aggregations, provides a mechanism linking PAR4
28 signaling to hypercoagulation and thromboinflammation. [5,42]

29
30 In the early stages of venous thrombosis, platelets are recruited to the site where they associate with
31 neutrophils and trigger TF expression, microparticle release, and NETosis [9,30]. This association is
32 initially mediated by platelet P-selectin and neutrophil PSGL-1, and further supported by activated
33 integrins on both cells [9,30]. Additionally, procoagulant platelets have characteristically high levels of
34 P-selectin on their surface, and are known to preferentially bind to neutrophils [5,29,37]. Previously, we
35 have shown that platelets from PAR4-322^{P/L} and PAR4-322^{L/L} mice require 2 and 8-fold higher
36 concentration of PAR4-AP (AYPGKF), respectively, than wild-types to express their maximum level of
37 P-selectin, and integrin activation was impaired to a similar degree [24]. Here, platelet-neutrophil
38 complexes were reduced concomitantly with decreased PAR4 reactivity (**Figure 7**). Complexes were
39 unable to form from PAR4-322^{L/L} mice even at 400 μ M, which triggers a full response from PAR4-322^{P/P}
40 (wild-type). Taken with our previous data, PAR4-mediated platelet-neutrophil interactions are
41 predominantly driven by PAR4 on platelets [24]. A direct role of PAR4 signaling on neutrophils cannot
42 be ruled out, however, in our hands we have not observed PAR4-mediated signaling on neutrophils.

43
44 Here we show that the PAR4-P322L mutation negatively impacts venous thrombosis and platelet
45 procoagulant activity. Our *ex vivo* experiments were consistent with previous observations that PAR4-
46 322^{L/L} platelets have a more dramatic phenotype than heterozygous PAR4-322^{P/L} platelets. This gene-
47 dosage effect implies that more leucines at PAR4-322 have a greater negative impact on platelet

1 response to PAR4 agonists like thrombin. Conversely, our data from IVC stasis experiments show that
2 the PAR4-322^{P/L} mice instead have a more dramatic phenotype than the PAR4-322^{L/L} mice, which are
3 more similar to wild-types. Previously, experiments in arterial thrombosis showed a similar trend where
4 PAR4-322^{P/L} mice, specifically females, had a more dramatic phenotype than their 322^{L/L} littermates
5 [24]. An important distinction is that this switch was only seen in females, and here we only use males
6 in our in vivo studies due to anatomical differences. It is also important to remember here that our
7 mutation is global, and it impacts PAR4 on endothelial cells as well. Recently, Rajala et al. have shown
8 that PAR4 has distinct signaling roles in liver endothelial cells [43]. We have only investigated the
9 impact of the mutation of platelets specifically in this process, and it is likely that the PAR4-P322L
10 mutation is also decreasing endothelial cell activation in response to thrombin.

11
12 PAR4 antagonists are currently not used therapeutically to prevent venous thrombosis. With our
13 growing knowledge of how platelets promote venous thromboembolism, it is worth exploring PAR4 as a
14 target. Venous clots are predominantly composed of red blood cells, fibrin, and neutrophils [32]. While
15 platelets do not make up a large volume of the final thrombus, these small cells are vital to the
16 beginning stages of thrombosis. Platelet participation in VTE has been known for over 50 years. In
17 1965, Hirsh and McBride reported that platelets from patients with recurrent VTE showed greater
18 adhesive properties [44]. Platelets drive VTE by binding to damaged endothelium where they activate
19 and promote thrombin generation and fibrin formation [30]. Here, we have shown that mice with the
20 PAR4-P322L mutation have reduced venous thrombi size and incidence. This recapitulates clinical
21 observations that humans with a leucine at PAR4-P310 are at a reduced risk of venous
22 thromboembolism. We have also directly associated the PAR4-P322L mutation with reduced platelet
23 procoagulant activity and complex formation with neutrophils, both of which promote thrombosis.
24 Importantly, reduced PAR4 activity did not have an impact on clot formation measured by ROTEM.
25 Combined with our data that reduced PAR4 function negatively impacts both arterial and venous
26 thrombosis, this highlights PAR4's therapeutic potential to reduce thrombosis without impairing
27 hemostasis.

28 29 **Author Contributions:**

30 Study Design: EK, ASG, MN Data Collection: EK, JC, NL, DD Data Analysis: EK, JC, NL, DD, MN
31 Drafting Manuscript: EK, MN Critical Revisions: JG, NL, GH, DD, ASG

32 33 **Authors conflict of interest statement:**

34 Elizabeth A. Knauss, Johana Guci, Norman Luc, Dante Disharoon, Grace H.Huang, Anirban Sen
35 Gupta, and Marvin T. Nieman all declare no conflicts of interest.

36 37 **Funding:**

38 Funding for this study was provided by the National Institutes of Health (NIH) National Heart Lung and
39 Blood Institute (NHLBI), HL098217 (MN), HL154026 (MN), HL121212 (ASG), and HL137695 (ASG),
40 the Ruth L. Kirschstein Predoctoral Individual National Research Service Award F31HL162548 (EK).
41 Histology services were supported by the Tissue Resources Shared Resource and flow cytometry was
42 supported by the Cytometry & Imaging Microscopy Shared Resource of the Case Comprehensive
43 Cancer Center (P30CA043703). This project was also supported by the Clinical and Translational
44 Science Collaborative of Northern Ohio which is funded by the National Institutes of Health, National
45 Center for Advancing Translational Sciences, Clinical and Translational Science Award grant,
46 UM1TR004528. The content is solely the responsibility of the authors and does not necessarily
47 represent the official views of the NIH

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1 **Figure Legends**

2 **Figure 1. PAR4-P322L Reduces Venous Thrombosis after Complete IVC Stasis.** (A) Venous clots
3 within the inferior vena cava (IVC) were harvested from wild-type, heterozygous PAR4-322^{P/L},
4 homozygous PAR4^{L/L}, and global PAR4^{-/-} mice 48 hours after complete IVC ligation. (B) Clot incidence
5 and (C) thrombus weight were also recorded 24 hours after IVC ligation. Dots represent individual mice.
6 Statistical significance was determined by unpaired t tests (A,C) and Fisher's exact test (B). **p<0.01
7

8 **Figure 2. PAR4-P322L Reduces Venous Thrombosis after Partial IVC Stenosis.**

9 (A) Thrombi were excised from the surrounding tissue and weighed 48 hours after IVC stenosis. Dots
10 represent individual mice. Statistical significance was determined by unpaired t tests *p<0.05
11

12 **Figure 3. PAR4-P322L Mice have Smaller Thrombi after IVC Stenosis.**

13 Venous clots were excised from the surrounding tissue, embedded in paraffin, and sectioned for (A)
14 haematoxylin and eosin (A and B) or carstairs (C and D) staining. Images were taken at 4x
15 magnification and stitched together when necessary to display the entire clot (panels A and C). The
16 stitching is indicated with the vertical dotted line. Higher resolution images (200x) are presented in
17 panels B and D.
18

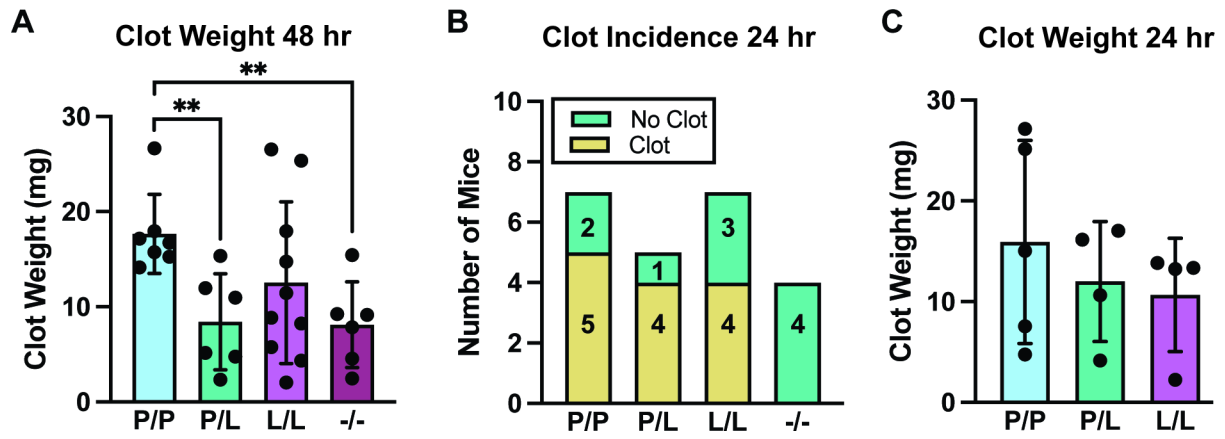
19 **Figure 4. Blood from PAR4^{-/-} Mice Shows Impaired Clot Formation.**

20 ROTEM was performed to measure coagulation parameters by generating a thromboelastogram. (A)
21 representative traces for each genotype are shown. (B) Clotting time, (C) clot formation time, (D)
22 maximum clot firmness, (E) alpha angle, and the amplitudes at (F) 10 and (G) 20 minutes were
23 extracted from the ROTEM traces. Dots represent individual mice. Statistical significance was
24 determined by unpaired t tests *p<0.05, ****p<0.0001.
25

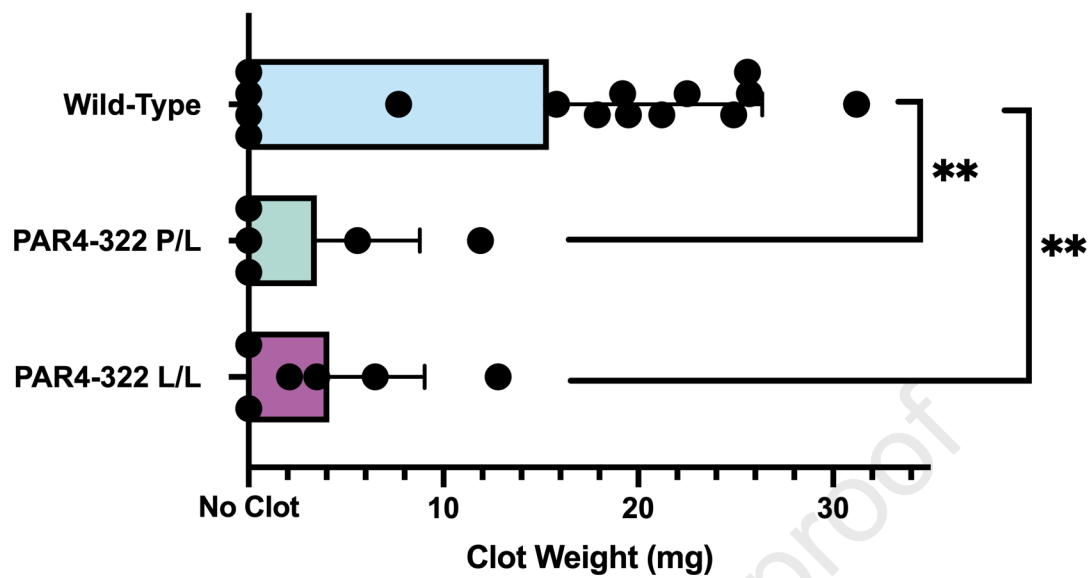
26 **Figure 5. PAR4-P322L Decreases Platelet-Mediated Thrombin Generation.** Platelet rich plasma
27 (PRP) was obtained from wild-type, PAR4-322^{P/L}, PAR4-322^{L/L}, and PAR4^{-/-} mice and assessed for
28 thrombin generating potential (A-D). (A) Thrombin generation curves were created using a fluorogenic
29 thrombin substrate over 1 hour. (B) The area under the curve for each genotype was compared to the
30 provided control human plasma (control high and control low) and interpreted as the total thrombin
31 generated over that hour. (C) The peak concentration of thrombin generated and (D) the time it took to
32 reach that peak was also recorded. Statistical significance was determined by unpaired t tests. *p<0.05
33

34 **Figure 6. PAR4-P322L Reduces Platelet Phosphatidylserine Surface Expression.** PS exposure
35 was measured with flow cytometry using FITC-tagged Lactadherin. Washed mouse platelets were
36 stimulated with convulxin (1 nM) in combination with 200 μM of the PAR4-AP (AYPGKF), 1 nM of
37 thrombin, and 5 nM of thrombin. Lactadherin-positive events were recorded as the percentage of
38 CD41-positive cells that were also positive for FITC. Dots represent individual mice. Statistical
39 significance was determined by unpaired t tests. *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001.
40

41 **Figure 7. PAR4-P322L Decreases Platelet-Neutrophil Aggregation.** Platelet-Neutrophil complexes
42 were measured using flow cytometry to record the percentage of LyG+ neutrophils that also expressed
43 CD41. Lysed whole blood from wild-type, PAR4-322^{P/L}, PAR4-322^{L/L}, and PAR4^{-/-} mice were incubated
44 with with (A) no agonist, or (B) 50 μM, (C) 200 μM, or (D) 400 μM of the PAR4-agonist peptide
45 (AYPGKF). Statistical significance was determined by unpaired t tests *p<0.05, **p<0.01, ***p<0.001,
46 ****p<0.0001.
47

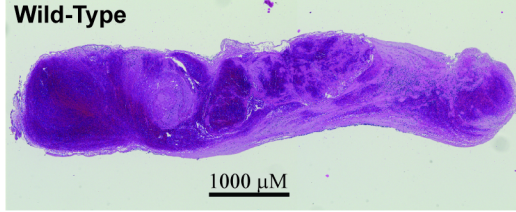


IVC Stenosis Weights at 48 hours

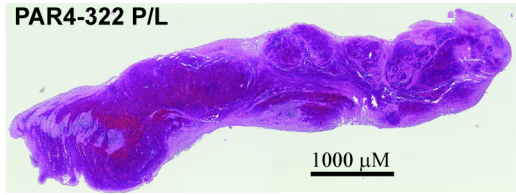


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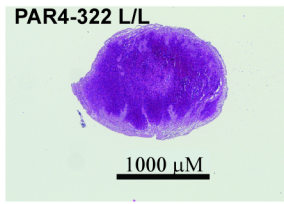
Wild-Type



PAR4-322 P/L

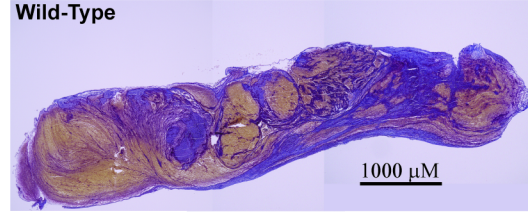


PAR4-322 L/L

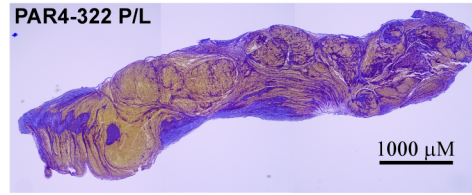


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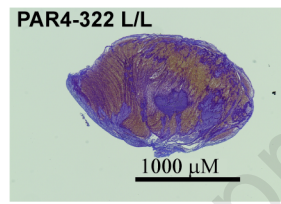
Wild-Type



PAR4-322 P/L



PAR4-322 L/L



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